



Meropenem-vaborbactam (20+10) M/V 30 µg

INTENDED PURPOSE

Meropenem-vaborbactam (20+10) M/V 30 µg is an *in vitro* semi-quantitative method for antimicrobial susceptibility of clinical isolates tested on agar media using overnight incubation.

The test is used against the following microorganisms:

Gram-negative bacteria

Enterobacterales

Pseudomonas aeruginosa

This device is for Research Use Only (RUO).

DESCRIPTION

Antibiotic Disc are paper discs with special features, that are impregnated with antibiotic and used for the susceptibility test according to the Kirby-Bauer antibiotic testing (KB testing or disk diffusion antibiotic sensitivity testing).

The simultaneous growth of the bacteria and diffusion of the antimicrobials compounds forms a zone of inhibition of growth. Zone size observed in a disk diffusion test has no meaning in and of itself. This information is correlated with *in vivo* test able to determinate resistance and susceptibility to antimicrobials and result in the interpretive standards.

Meropenem-vaborbactam (20+10) M/V 30 µg Disc is used for determining bacterial antibiotic susceptibility in the treatment of infectious disease.

Meropenem-vaborbactam concentration is 20 µg of meropenem and 10 µg of vaborbactam.

KIT CONTENT

Antibiotic Disc is supplied in different packaging options (no additional reagents are included):

Disc in cartridge

- The 50-test box contains 1 cartridge with 50 discs packed in desiccant envelopes.
- The 250-test box contains 5 cartridges of 50 discs, each cartridge individually packed in a desiccant envelope.
- Each package also contains a transparent resealable bag.

Discs in canister

- The canister contains 250 discs and a desiccant tablet.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as:

- sterile loops, swabs (not too tightly spun), test tubes, pipettes and scissors
- suspension medium - McFarland turbidity standard - agar plate medium (validated by the media manufacturer for use with antimicrobial susceptibility testing, 90- or 150-mm plates)
- forceps - incubator - quality control organisms

PRINCIPLE OF THE METHOD

The discs are applied to the surface of a culture medium inoculated with a pure colony suspension of the microorganism under examination. After incubation, the plates are examined, the inhibition zone around each disc are examined and compared with the standard disc diffusion interpretive criteria: in this way the microorganisms are defined as being susceptible, intermediate or resistant to the tested antimicrobial agents.

SPECIMEN COLLECTION AND PREPARATION

Antibiotic Discs are not for use directly with clinical or other specimens. The product is used to indicate appropriate patient treatment against infections caused by microorganisms that can be isolated from clinical samples of adult, juvenile and pediatric patients. There are no different indications for use according to sample source.

The microorganism to be tested must first be isolated on a nonselective culture medium, such as blood agar or tryptic soy agar (TSA). In case of mixed culture, selected colonies should be purified by subculturing. Differential media harboring chromogenic or fluorogenic substrates should not be used for the subculture. It is recommended that cultures be no more than 24 hours old unless additional incubation is required to achieve sufficient growth.

TEST PROCEDURE

Handling

Before using the Antibiotic Disc from an unopened package, visually inspect to ensure the package is intact. Do not use the discs if the package has been damaged. When removed from the refrigerator/freezer, allow the package or storage container to reach room temperature for about 30 minutes. Moisture condensing on the outer surface must evaporate completely before opening the package. Use forceps or a similar device to pick up a disc.

When using Antibiotic Discs from a canister, replace the lid immediately after use and store as outlined under STORAGE.

Inoculum Preparation

Suspend well-isolated colonies from an overnight agar plate into the suspension medium to achieve the recommended McFarland standard. If the inoculum concentration is correct, a confluent lawn of growth will be obtained after incubation. If insufficient growth occurs, the testing should be repeated.

McFarland turbidity standards do not guarantee the correct number of viable cells in the suspension. In order to verify that your procedure gives the correct inoculum density in terms of CFU/mL performing regular colony counts is recommended. An acceptable inoculum should give approximately $1-2 \times 10^8$ CFU/mL.

Inoculation

Dip a sterile swab in the broth culture or in a diluted form thereof and squeeze it on the wall of the test tube to eliminate excess liquid. Streak the swab over the entire sterile agar surface. Repeat this procedure by streaking 2 more times, rotating the plate approximately 60 degrees each time to ensure an even distribution of inoculum. Allow excess moisture to be absorbed so that the surface is completely dry before applying discs.

Use well-defined, high-quality media for AST that supports good growth. The brand chosen should have good batch-to-batch reproducibility to ensure that accurate and reliable MIC values are obtained.

The agar medium should have a depth of 4.0 ± 0.5 mm, a pH of 7.3 ± 0.1 and all other quality specifications should be fulfilled. Refer to the media manufacturer's instructions for more information.

Application

Apply the disc to the inoculated agar surface. Pressing it with a sterile forceps on the surface of the agar. Once applied, do not move the disc.

Incubation

Incubate the agar plates in an inverted position at the appropriate temperature, atmosphere and time, according to the methodology followed.

NOTES:

- The medium to be used depends on the organism under investigation and the methodology followed and must be validated by the media manufacturer for antimicrobial susceptibility testing.
- It is recommended to use the inoculum suspension within 15 minutes of preparation, apply discs within 15 minutes of inoculation and incubate plates within 15 minutes of disc application.

For more details, please refer to the current published standards.

READING THE RESULTS

At the end of the incubation period, measure the inhibition zone diameters (mm) with zone edges should be read at the point of complete inhibition as judged by the naked eye with the plate held about 30 cm from the eye.

Measure zone diameters to the nearest millimeter with a ruler or a calliper. If an automated zone reader is used, it must be calibrated to manual reading.

Do not read the plate if the culture appears mixed or if the lawn of growth is too light or too heavy.

For full instructions relating to the interpretation of the results according to CLSI and/or EUCAST methodology please refer to the relevant current standards.

NOTES:

- Excessively wet plates prior to inoculation, insufficient drying before applying discs and/or unevenly streaked surfaces may give non confluent growth. Repeat the test if the inhibition zone diameter is difficult to read.
- Occasionally, certain antimicrobial agent/microorganism combinations may give unusual results. In these cases, judgment of the inhibition zone diameter may be difficult for the inexperienced personnel.

Procedures specific to Meropenem-vaborbactam (20+10) M/V 30 µg Disc are summarized in the following table:

Storage	Temperature at $-20^{\circ}\text{C} \pm 5^{\circ}\text{C}$
Organism	Enterobacterales, <i>Pseudomonas aeruginosa</i>
Medium	Mueller Hinton II Agar
Inoculum	Suspension in saline (0.85% NaCl) to 0.5 McFarland standard (1 if mucoid)
Incubation	Agar plates in inverted position at $35 \pm 2^{\circ}\text{C}$ for 16-18 h in ambient air (CLSI) and $35 \pm 1^{\circ}\text{C}$ in air for 18 ± 2 h (EUCAST)

INTERPRETATION OF THE RESULTS

To categorize the result, typically as susceptible, intermediate or resistant, refer to current Antibiotic Disc breakpoints (below).

The inhibition zone diameters obtained should be interpreted according to current CLSI and EUCAST interpretive criteria (see below).

Antimicrobial agent	Organisms	CLSI		
		Zone diameter breakpoint (mm)		
		S $\geq$	I	R $\leq$
Meropenem-vaborbactam	Enterobacterales (excluding <i>Salmonella</i> and <i>Shigella</i> spp.)	18	15-17	14

Antimicrobial agent	Organisms	EUCAST		
		Zone diameter breakpoint (mm)		
		S $\geq$	R <	ATU
Meropenem-vaborbactam	Enterobacterales	20	20	15-19
	<i>P. aeruginosa</i>	14	14	

Disclaimer: This breakpoint table might be out-of-date and does not replace CLSI and EUCAST published guidelines, which always should be consulted before disc categorization.

USER QUALITY CONTROL

To check the performance of Antibiotic Disc reagents, media and procedure, test the quality control strain(s) as shown below. Results are considered satisfactory if the quality control result(s) fall within the expected range(s).

Antimicrobial agent	Control strain		Zone Range (mm)
Meropenem-vaborbactam M/V	<i>Escherichia coli</i>	ATCC® 25922	31 – 37 ^{a,b}
	<i>Pseudomonas aeruginosa</i>	ATCC® 27853	29 – 35 ^{a,b}
	<i>Klebsiella pneumoniae</i>	ATCC® 700603	29 – 35 ^a
	<i>Klebsiella pneumoniae</i>	ATCC® BAA-1705	21 – 27 ^a
	<i>Klebsiella pneumoniae</i>	ATCC® BAA-2814	16 – 20 ^a
	<i>Klebsiella pneumoniae</i>	ATCC® BAA-2814	15– 21 ^b

a) CLSI M100-Ed36

b) EUCAST Quality Control v.16.0

LIMITATIONS

Invalid results can be caused by poor specimen quality, improper sample collection, improper transportation, improper laboratory processing, or a limitation of the testing technology. The operator should understand the principles of the procedures, including its performance limitations, in advance of operation to avoid potential mistakes.

WARNINGS AND PRECAUTIONS

- 1) **For Research Use Only (RUO).**
- 2) **For laboratory professional use only.**
- 3) **The disc is for single use only and should not be reused.**
- 4) Operators must be trained and have certain experience. Please read the instructions carefully before using the kit. Reliability of assay results cannot be guaranteed if there are any deviations from the instructions in this document.
- 5) Do not use if material from a packaging or the packaging itself appear to be damaged.
- 6) Avoid cross-contamination of samples by using disposable tips and changing them after each sample.
- 7) Do not mix reagents of different batches. Please use the kit within the validity period.
- 8) Do not eat, drink, smoke, apply cosmetics or handle contact lenses in areas where reagents and human specimens are handled
- 9) Ensure laboratory equipment is calibrated and maintained in accordance with the laboratory's procedure.

STORAGE

Unopened foil packages and canisters: On receipt, store Antibiotic Disc at –20°C to +8°C until the given expiry date. Some Antibiotic Disc (e.g. carbapenems) should be stored frozen at –20°C ± 5°C. Check the drug label for the specific storage temperature.

Open foil packaging: Leftover discs from an opened CARTRIDGE need to be stored at 2-8°C for no more than 7 days. The cartridge containing unused discs should be returned into its desiccant envelope and then inserted into the resealable bag.

Opened canisters: Discs in canister can be used for up to 2 months from first opening (record the date on which the canister was open) and must be stored at the label storage temperature. Before using the remaining discs, check the expiry date indicated on the packaging. Do not store near sources of heat and do not expose to excessive temperature variations.

Protect Antibiotic Disc from moisture, heat and direct exposure to strong light at all times.

DISPOSAL OF USED MATERIAL

After use, the discs and material that has come into contact with the sample must be decontaminated and disposed of in accordance with guidelines used in the laboratory for decontamination and disposal of potentially infected material.

SUGGESTIONS FOR TROUBLESHOOTING

For out-of-range QC, first repeat the test with a pure culture or a freshly subcultured QC strain. If the issue is unresolved, follow this guidance for additional suggestions for troubleshooting out-of-range QC results and unusual clinical isolates results.

Observation	Probable Cause	Comments/Suggested Actions
Inhibition zone diameter too large	Inoculum too light	Repeat using McFarland 0.5 turbidity standard or standardizing device. Check expiration date and proper storage if using barium sulfate or latex standards. Check steps in inoculum preparation and incubation procedure. Perform colony count check of growth control well immediately after inoculation and before incubation (<i>E. coli</i> ATCC® 25922 closely approximates 5 x 10 ⁵ CFU/ml)
Inhibition zone diameter too small	Inoculum too heavy	
Inhibition zone diameter too small	Antimicrobial agent is degrading	Use alternative lot. Check STORAGE and package integrity

REFERENCES

1. CLSI. Performance Standards for Antimicrobial Susceptibility Testing. 36th ed. CLSI supplement M100. Clinical and Laboratory Standards Institute; 2026.
2. CLSI. Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. 12th ed. CLSI standard M07. Wayne, PA: Clinical and Laboratory Standards Institute; 2024
3. CLSI. Verification of Commercial Microbial Identification and Antimicrobial Susceptibility Testing Systems. 1st ed. CLSI guideline M52. Wayne, PA: Clinical and Laboratory Standards Institute; 2015.
4. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and diameters zones. Version 16.0, 2026. <http://www.eucast.org>.
5. The European Committee on Antimicrobial Susceptibility Testing. Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST. Version 16.0, 2026. <http://www.eucast.org>
6. ISO 20776-1:2019. Clinical laboratory testing and in vitro diagnostic test systems -- Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices -- Part 1: Reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious disease.
7. ISO 20776-2:2021. Clinical laboratory testing and in vitro diagnostic test systems - Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices -- Part 2: Evaluation of performance of antimicrobial susceptibility test devices

Product	µg	Code	Packaging	Ref.
Meropenem-vaborbactam	(20+10) 30	M/V	5 x 50	9326
			1 x 50	9326/1
			1 x 250	9326/2

Table of Symbols

	For Research Use Only
	Catalogue number
	Batch code
	Do not reuse
	Fragile, handle with care
	Manufacturer
	Use by
	Contains sufficient for <n> tests
	Consult instructions for use
	Temperature limits

Revision History

Revision	Release Date	Change Summary
0	11 May 2026	Document creation

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CLSI is a trademark belonging to Clinical Laboratory and Standards Institute, Inc.

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EUCAST stands for European Committee on Antimicrobial Susceptibility Testing. These data have been made available at no cost by EUCAST and can be accessed freely on the EUCAST website: www.eucast.org. EUCAST recommendations are frequently updated, and the latest versions are available at www.eucast.org.

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